

Methods

Sample collection and preparation

Oragene•Dx saliva samples were collected from 27 healthy donors. These samples were pre-screened for DNA yield by using prepIT•Q2A preparation reagent followed by PicoGreen™ fluorescence quantification. Seven of these samples were selected to cover the range of DNA yield observed in a typical population, as well as emphasize donors with high DNA content to push the limits of the direct-to-library capability of this workflow (Figure 1). DNA was prepared from the selected samples in duplicate using prepIT•Q2A. Purified genomic DNA from cell line NA12878 (Coriell Institute) was normalized to 5ng/μL (50 ng total) and was used as a control in duplicate.

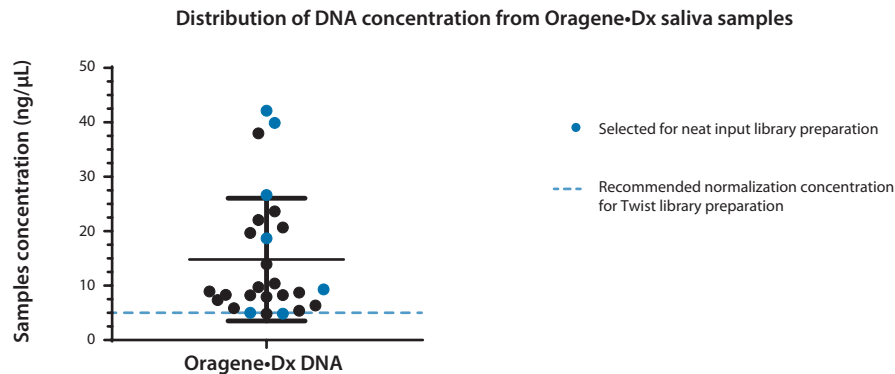


Figure 1: Distribution of DNA concentration from Oragene•Dx saliva in a healthy population and concentration of selected samples for library preparation.

Twist library preparation

Samples were prepared according to Enzymatic Fragmentation and Universal Adapter System protocol with the following modifications. The standard protocol requires samples normalized to 5 ng/μL with a 10 μL input volume, while in this study, 10 μL neat was used directly in the workflow without normalization. In addition, fragmentation time was reduced to 18 and 12 minutes to ensure that the correct target fragment size was achieved. The Agilent Bioanalyzer DNA 7500 assay was used to confirm fragment length, as shown in Figure 2, and all samples achieved the target fragment size despite the sample input amount used. Hybridization steps were performed as written in the protocol.

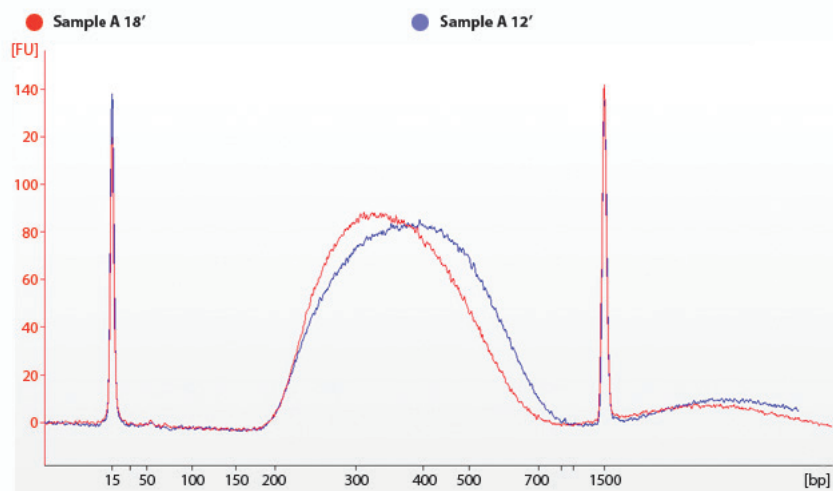
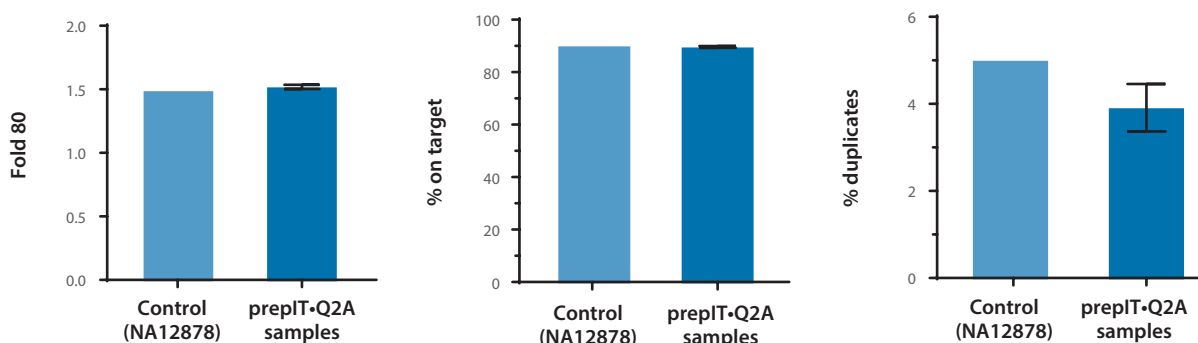


Figure 2: Tunability using enzymatic fragmentation. At 18 minutes the average fragment size was 368 bp. At 12 minutes the average size was 393 bp. The concentrations were 161 ng/μL and 184 ng/μL respectively. Shown here with a 3x dilution.

Following library preparation, samples (N=8 including control) were pooled together for a multiplex capture reaction. The Twist Human Core Exome Panel was used with Fast Hybridization Protocol with a hybridization time of two hours. The enriched libraries were sequenced on the NextSeq 500/550 High Output v2 kit (Illumina®) to generate 2x76 paired-end reads and down-sampled to 150x of targeted bases. Picard HS_metrics tools with a mapping quality of 20 were utilized for sequence analysis.

The quality of the capture reaction was evaluated using Picard metrics¹. prepIT•Q2A samples had consistent performance with the control NA12878 as demonstrated in Figure 3 by similarly high on target rates (> 90 %), low duplication rates (< 5 %), and high uniformity (fold 80 < 1.5). In addition, there was no relationship between sample input amount and quality metrics, indicating that this workflow is robust and compatible with direct prepIT•Q2A sample input into library preparation without impact on downstream performance.



Conclusion

A workflow for Oragene•Dx using prepIT•Q2A with Twist NGS library preparation and target enrichment components was presented. The workflow demonstrated the compatibility of saliva prepared with prepIT•Q2A direct into Twist NGS library preparation and exome sequencing, with no need to quantify or normalize DNA samples. The results illustrate the robust performance with high on-target rates, low duplication rates, and high uniformity across samples. Further to this, the workflow is streamlined and enables sample to sequencer within a standard 8-hour work day.

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